

## Effect of Chromium and Selenium-E on Progesterone and Estradiol-17 $\beta$ Levels during Reproductive Cycle of Baladi Female Goats under Egyptian Conditions

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### **ABSTRACT**

The effect of chromium and selenium with vitamin E on progesterone ( $P_4$ ) and estradiol-17  $\beta$  ( $E_2$ ) levels of Baladi female goats was the objective of this study as new techniques for improving the reproductive performance of Egyptian goats. Seventy-two of mature Baladi does were used through two conditions of mild and hot climates (36 animals / season). The animals were randomly divided into three equal groups, the 1<sup>st</sup> is the control group, the 2<sup>nd</sup> group was supplemented by chromium in capsules and the 3<sup>rd</sup> group was intramuscularly injected twice a week with 2ml VITSELEN®, (0.5 mg selenium and 10.7 IU vitamin E, Se-E). Serum hormones levels were assessed in the three experimental goats at the estrous, pregnancy and postpartum periods. The results showed that  $P_4$  levels during estrous cycle and mid-pregnancy periods were significantly higher in mild climate than in hot climate while opposite trend was found during early and late pregnancy as well as postpartum period.  $E_2$  levels during postpartum period were significantly higher in hot climate than in mild while during estrous cycle no significant differences in  $E_2$  levels due to season. Chromium showed significant ( $P < 0.05$ ) increase in  $P_4$  levels in goats during each of estrous cycle and pregnancy periods and showed significant decrease in  $P_4$  levels in goats during postpartum period as compared with control. Selenium-E significantly increased the concentration of  $P_4$  in goats during each of the luteal phase of estrous cycle, pregnancy and at 30<sup>th</sup> day postpartum periods as compared with control. The opposite trend was found at 15<sup>th</sup> and 45<sup>th</sup> days postpartum. No significant difference

### **KEYWORDS**

*Chromium,  
Selenium-E, Heat  
Stress, Progesterone,  
Estradiol-17 $\beta$ , Goats.*

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in P4 due to Selenium-E during follicular phase of estrous cycle. Chromium showed significant decrease in E<sub>2</sub> levels in goats on 15<sup>th</sup> and 45<sup>th</sup> days postpartum period while no significant differences in E<sub>2</sub> levels due to chromium treatment during estrous cycle and 30<sup>th</sup> day of postpartum period. Selenium-E increased significantly E<sub>2</sub> level during estrous cycle and postpartum periods.

## INTRODUCTION

**H**eat stress has a reducing effect on both the reproductive and productive efficiency of farm animals. It has been shown to alter the duration of estrus, conception rate, endocrine status, follicular growth and development; early embryonic development and fetal growth (Jordan, 2003). The breeding season of goats begins when the photoperiod become shorter, depending on daylight, breed and diet (Chemineau et al. 1999). In Egypt, as the days begin to shorten in the late summer and early autumn, a regular estrus cycle is initiated; and in late winter and early spring when the days are lengthening; partial anestrus occurs. Egyptian Baladi goats shows normal breeding season at mid-summer and continue through the autumn season. However, a considerable percentage of Baladi goats exhibit estrus signs during out-of-season period (32%) (El-Tarabany et al., 2017 & 2018).

In goats, heat stress reduced plasma concentrations of estradiol and lowered follicular estradiol concentration, LH receptor level, delayed ovulation (Ozawa et al., 2005), altered follicular growth (Roth et al., 2000 and Hansen, 2009), and disrupt the development and function of the oocytes (AL-Katanani et al., 2002; Sartori et al., 2002 and Hansen, 2009). Roth et al. (2000) suggested that early atresia in medium sized follicles because of heat stress could be associated with low estradiol production by granulosa cells and increased progesterone concentrations in the fol-

licular fluid of heat-stressed cows. In addition, heat stress can cause increased cortisol secretion (Wise et al., 1988 and Elvinger et al., 1992), and ACTH has been reported to block estradiol-induced sexual behavior (Hein and Allrich, 1992). Emesih et al. (1995) reported that pregnant doe goats exposed to heat stress showed higher plasma progesterone concentrations than in those maintained at moderate ambient temperature.

Studying chromium importance as an essential element for livestock animals intensively started in 1990s (Pechova and Pavlata 2007). Garcia et al. (1997) have studied the effect of Cr-picolinate supplementation on sensitivity of tissues to insulin, ovulation rate, and progesterone and oxytocin secretion; Cr supplementation had a positive effect on the ovulation rate but progesterone concentrations remained unchanged. A positive effect of Cr supplementation on the insemination index, interval and service period has been established (Bonomi et al., 1997 and Pechova et al., 2003). Selenium, an essential part of the antioxidant defense system, plays an important role in the growth and health of humans and livestock through its participation in several important enzymes and enzyme reactions (Surai, 2006). Selenium deficiency has been related to reproductive failures in ruminants, and Vitamin-E is an important complementary factor (Buchanan-Smith et al., 1969).

The issues of reproduction in relation to Cr supplementation have been devoted relatively little attention, so this paper aims to find out the effect of chromium and selenium-E administration on progesterone and estradiol-17 $\beta$  concentrations during estrous cycle, pregnancy and postpartum periods of Baladi doe goats under Egyptian conditions.

## MATERIALS AND METHODS

### *Animals and feeding:*

This experiment was carried out in Goats Experi-

mental Farm, Nuclear Research Center, Atomic Energy Authority, Inshas; (latitude 31° 12' N to 22° 2' N, longitude 25° 53' E to 35° 53' E). The experimental animals were healthy and clinically free of external and internal parasites and were fed basal ration of concentrate feed mixture (CFM) according to the allowances of NRC (2007) of goats. The CFM composed of 37.4% wheat bran, 27% yellow corn, 12.5% soybean meal, 10.0% undecorticated cottonseed cake, 5% rice bran, 4% sugarcane molasses, 3% limestone, 1% sodium chloride and 0.1 vitamin and minerals premix. Feed mixture was offered once daily at 9:00 a.m., based on 3.5% of body weight. Barseem hay was offered *ad libitum*. Fresh drinking water was available at all time.

#### **Animal housing:**

All experimental animals were kept in semi-open pens throughout the experimental period. These pens were provided with enough shade and ventilation in summer and protection from rain in winter. The does were allowed to graze five hours daily at least.

#### **Experimental design:**

Seventy-two mature female goats (36 animals/ season) aged 2-3 years old with average body weigh  $25 \pm 1.5$  kg were randomly divided into three equal groups. Animals in the 1<sup>st</sup> group were kept as control, the 2<sup>nd</sup> group was supplemented by chromium (chromium chloride trivalent), 0.8 mg/head/day as capsules (Williams *et al.*, 1994) and the 3<sup>rd</sup> group was intramuscularly injected with 2 ml VITSELEN®, contained 0.5 mg selenium and 10.7 IU vitamin E/head/day.

#### **Estrus synchronization and blood sampling:**

All groups received 10 ml of PGF2 $\alpha$  (lutalyse) in double dose (5 mg/ dose, IM) at 11 day-intervals, followed by 500IU of hCG; then after 24 hours three fertile bucks (one buck for each group) were introduced to the does and allowed to be with does for two successive estrous cycles for estrous detection and natural mating. Blood samples were collected from the jugular vein in evacuated glass tubes, and kept at room temperature from 30 to 60 min for clotting, then centrifuged at 3000 rpm for 15 min to separate serum. After that serum was stored at -20°C until analyzed. Samples were collected throughout different stages of estrous cycle, according to Fatet *et al.* (2011); monthly during pregnancy and every 15-day up to 45 days postpartum.

#### **Ambient temperature, relative humidity and temperature humidity index:**

The ambient temperature and relative humidity were recorded daily from meteorological station of Atomic Energy Authority during the whole experimental period. The temperature humidity index (THI) was calculated during mild and hot climates according to Marai *et al.* (2000) as:

$THI = db\ ^\circ C - [(0.31 - 0.31RH) \times (db\ ^\circ C - 14.4)]$   
where, THI= temperature humidity index, db °C= dry bulb temperature in Celsius and RH = relative humidity 100%. A value for THI < 22.2 was considered remarkably an absence of heat stress, while the values from 22.2 to 23.3 referred to moderate (mild) heat stress, 23.3 to <25.6 referred to severe heat stress and >25.6 referred to very severe heat stress.

**Table (1) : Mean Values of THI during the experimental period.**

Climate	Ambient temperature		Relative humidity values %		THI	
	Max	Mini	Max	Mini	Max	Mini
Mild	23.47	14.24	80.01	27.44	22.9	13.9
Hot	34.15	23.28	77.91	20.30	32.8	23.5

Mild= October, November, December, January, February and March; Hot= May, June, July, Augustus, September, October.

**Progesterone and estradiol-17 $\beta$  (P<sub>4</sub> and E<sub>2</sub>):**

Serum P<sub>4</sub> (ng/ml) and E<sub>2</sub>-17 $\beta$  (Pg/ml) concentrations were determined using Coat-A-count I<sup>125</sup> RIA kits obtained from Immunotech a Beckman Coulter Company, Czech Republic.

**Statistical analysis:**

Data were expressed as mean  $\pm$ SE. Data were analyzed statistically by GLM procedure of the SAS program (SAS, 1998) according this equation:

$Y_{ijk} = \mu + S_i + T_j + (ST)_{ij} + e_{ijk}$  where,  $Y_{ijk}$  = the dependent variables estimated,  $\mu$  = Overall mean,  $S_i$  = the effect of  $i^{\text{th}}$  season (1=mild and 2=hot),  $T_j$  = the effect of  $j^{\text{th}}$  treatment (1=control, 2=chromium and 3=selenium-E),  $(ST)_{ij}$  = the effect of interaction between season and treatment and  $e_{ijk}$  = random error.

Duncan's Multiple Range test was used to detect the differences among means of the experimental groups (Duncan, 1955).

**RESULTS****Effect of climate conditions and treatments on progesterone (P<sub>4</sub>) levels.**

Inspection the effect of treatments on P<sub>4</sub> concentrations, it is noticeable that P<sub>4</sub> levels were significantly higher in mild climate than in hot with the highest value 19.53 ng/ml at luteal phase in mild climate versus 11.21 ng/ml in hot one. Cr supplementation resulted in significant increase in (P<0.05) P<sub>4</sub> level during estrous cycle compared with other groups. The response to Cr treatment found to be a higher magnitude (25.39 ng/ml) during luteal phase, followed by Se-E (13.11ng/ml). Se-E does have higher (P<0.05) P<sub>4</sub> concentration at luteal phase than control, with values of 11.79 ng/ml vs. 6.57 ng/ml, respectively (Table 2).

Under mild climate conditions, treated groups

were higher (P<0.05) in P<sub>4</sub> than control during estrous cycle. Does of Cr group had the highest in P<sub>4</sub> levels (40.40 ng/ml) at luteal phase, followed by Se-E (13.86 ng/ml). On the other side, under hot climate, Cr or Se-E supplementations had no significant effect on P<sub>4</sub> concentrations as compared to control at two the phases of estrous cycle. Results obtained revealed that serum P<sub>4</sub> levels significantly (P<0.05) affected by climate, treatment and their interactions during pregnancy (Table 2).

Treating does by Cr or Se-E increased (P<0.05) P<sub>4</sub> concentrations during early, mid- and late-pregnancy more than control. Cr treatment almost had P<sub>4</sub> concentrations higher than Se-E, the highest value of 45.55 ng/ml was at mid-pregnancy, followed by 31.22 ng/ml for Se-E versus 21.72 ng/ml for control. During early and late-pregnancy, there was no significant difference between Cr and Se-E groups in P<sub>4</sub> levels. Marked increase in P<sub>4</sub> concentrations was found due to Cr and Se-E treatments with mean values of 62.95 and 39.61ng/ml, respectively, at mid-pregnancy of mild climate. The significant effect due to Cr or Se-E treatments under hot climate conditions was an increase (21.17 ng/ml, P<0.05) at early pregnancy and a decrease (21.24 ng/ml) at late pregnancy comparing with control, otherwise the treatments did not show significant differences in P<sub>4</sub> levels during pregnancy period (Table 2).

During postpartum period Cr treatment showed significant decrease (P<0.05) in P<sub>4</sub> concentration on 15<sup>th</sup> and 54<sup>th</sup> as compared with control especially under hot climate conditions. On the other side, Se-E resulted in an increase (5.05ng/ml; P<0.05) in P<sub>4</sub> concentration on day-30 PP comparing with other groups. This increase was clearly shown during hot climate with mean values 9.36ng/ml vs. 1.74 and 1.17ng/ml for Se-E, Cr and control, respectively. Otherwise, both treatments were lower than control (Table 2).

**Table (2) :** Effect of chromium and selenium-E on progesterone levels (ng/ml; means  $\pm$ SE) during estrous, pregnancy and postpartum periods of Baladi does.

Items	Progesterone (ng/ml; means $\pm$ SE)							
	Estrous cycle		Pregnancy period			Postpartum period		
	Follicular Phase	Luteal phase	Early	Mid	Late	15-day PP	30-day PP	45-day PP
<b>Climate condition (s)</b>								
<b>Mild</b>	2.10 $\pm$ 0.19	19.53 $\pm$ 2.71	9.72 $\pm$ 0.6	39.24 $\pm$ 4.41	22.22 $\pm$ 1.9	0.95 $\pm$ 0.04	0.61 $\pm$ 0.02	0.38 $\pm$ 0.02
<b>Hot</b>	1.62 $\pm$ 0.05	11.21 $\pm$ 0.56	18.88 $\pm$ 0.8	26.42 $\pm$ 1.5	25.17 $\pm$ 1.1	2.15 $\pm$ 0.12	4.42 $\pm$ 0.62	1.87 $\pm$ 0.16
<b>P- value</b>	0.001	0.0001	0.0001	0.0001	0.069	0.0001	0.0001	0.0001
<b>Treatments</b>								
<b>Control</b>	1.66 <sup>B</sup> $\pm$ 0.09	7.60 <sup>C</sup> $\pm$ 0.75	12.02 <sup>B</sup> $\pm$ 1.2	21.72 <sup>C</sup> $\pm$ 1.6	18.56 <sup>B</sup> $\pm$ 2.2	1.77 <sup>A</sup> $\pm$ 0.19	1.35 <sup>B</sup> $\pm$ 0.17	1.85 <sup>A</sup> $\pm$ 0.29
<b>Cr</b>	2.39 <sup>A</sup> $\pm$ 0.30	25.39 <sup>A</sup> $\pm$ 3.40	15.09 <sup>A</sup> $\pm$ 1.5	45.55 <sup>A</sup> $\pm$ 6.1	27.09 <sup>A</sup> $\pm$ 1.5	1.27 <sup>C</sup> $\pm$ 0.07	1.14 <sup>B</sup> $\pm$ 0.13	0.73 <sup>C</sup> $\pm$ 0.08
<b>Se-E</b>	1.53 <sup>B</sup> $\pm$ 0.07	13.11 <sup>B</sup> $\pm$ 0.68	15.80 <sup>A</sup> $\pm$ 1.0	31.22 <sup>B</sup> $\pm$ 1.9	25.43 <sup>A</sup> $\pm$ 1.6	1.61 <sup>B</sup> $\pm$ 0.17	5.05 <sup>A</sup> $\pm$ 0.95	0.79 <sup>B</sup> $\pm$ 0.1
<b>P- value</b>	0.0001	0.0001	0.003	0.0001	0.0001	0.0001	0.0001	0.0001
<b>Climate x Treatments interactions</b>								
<b>Mild</b>								
<b>Control</b>	1.65 <sup>b</sup> $\pm$ 0.18	4.32 <sup>c</sup> $\pm$ 0.06	7.01 <sup>d</sup> $\pm$ 0.3	15.15 <sup>d</sup> $\pm$ 0.2	9.76 <sup>c</sup> $\pm$ 1.3	0.88 <sup>d</sup> $\pm$ 0.05	0.53 <sup>c</sup> $\pm$ 0.03	0.46 <sup>d</sup> $\pm$ 0.016
<b>Cr</b>	3.35 <sup>a</sup> $\pm$ 0.33	40.40 <sup>a</sup> $\pm$ 2.49	9.00 <sup>d</sup> $\pm$ 0.5	62.95 <sup>a</sup> $\pm$ 8.9	27.29 <sup>a</sup> $\pm$ 2.8	1.16 <sup>c</sup> $\pm$ 0.08	0.55 <sup>c</sup> $\pm$ 0.02	0.33 <sup>e</sup> $\pm$ 0.012
<b>Se-E</b>	1.30 <sup>b</sup> $\pm$ 0.01	13.86 <sup>b</sup> $\pm$ 0.58	13.16 <sup>c</sup> $\pm$ 1.3	39.61 <sup>b</sup> $\pm$ 1.5	29.63 <sup>a</sup> $\pm$ 2.4	0.80 <sup>d</sup> $\pm$ 0.02	0.75 <sup>c</sup> $\pm$ 0.04	0.33 <sup>e</sup> $\pm$ 0.01
<b>Hot</b>								
<b>Control</b>	1.66 <sup>b</sup> $\pm$ 0.10	10.89 <sup>b</sup> $\pm$ 0.63	17.03 <sup>b</sup> $\pm$ 1.3	28.29 <sup>bc</sup> $\pm$ 1.5	27.36 <sup>a</sup> $\pm$ 2.5	2.65 <sup>a</sup> $\pm$ 0.08	2.17 <sup>b</sup> $\pm$ 0.10	3.25 <sup>a</sup> $\pm$ 0.01
<b>Cr</b>	1.44 <sup>b</sup> $\pm$ 0.05	10.38 <sup>b</sup> $\pm$ 0.94	21.17 <sup>a</sup> $\pm$ 1.6	28.14 <sup>bc</sup> $\pm$ 3.4	26.91 <sup>a</sup> $\pm$ 1.3	1.38 <sup>c</sup> $\pm$ 0.12	1.74 <sup>b</sup> $\pm$ 0.03	1.14 <sup>c</sup> $\pm$ 0.05
<b>Se-E</b>	1.76 <sup>b</sup> $\pm$ 0.11	12.36 <sup>b</sup> $\pm$ 1.22	18.45 <sup>ab</sup> $\pm$ 1.1	22.82 <sup>cd</sup> $\pm$ 0.7	21.24 <sup>b</sup> $\pm$ 1.1	2.42 <sup>b</sup> $\pm$ 0.08	9.36 <sup>a</sup> $\pm$ 0.63	1.23 <sup>b</sup> $\pm$ 0.02
<b>P- value</b>	0.0001	0.0001	0.01	0.0001	0.0001	0.0001	0.0001	0.0001

Means with different letters (A, B and C or a, b, c, ...) in the same column are significantly different at ( $P < 0.05$ ).

**Effect of climate conditions and treatments on estradiol-17 $\beta$  (E<sub>2</sub>) levels.**

It is obviously clear that, female goats treated with Cr or Se-E were higher in E<sub>2</sub>-17 $\beta$  concentrations more than control throughout follicular and luteal phases of estrous cycle, but the significant (P<0.05) increase was due to Se-E injection with mean value of 108.56 pg/ml during follicular phase (Table 3).

Under mild conditions, the significant (P<0.05) increase in E<sub>2</sub>-17 $\beta$  was 134.23 and 39.72pg/ml during follicular and luteal phases, respectively due to Se-E supplementation. However, Cr increased E<sub>2</sub>-17 $\beta$  concentrations more than control throughout estrous cycle of mild or hot climate conditions. This increase was significant (P<0.05) at luteal phase of hot season with mean value of 31.52pg/ml vs. 26.06 pg/ml for control (Table 3).

**Table (3) :** Effect of chromium and selenium-E on Estradiol-17 $\beta$  levels (pg/ml means  $\pm$ SE) during estrous and postpartum periods of Baladi does.

Items	Estradiol-17 $\beta$ (pg/ml; means $\pm$ SE)				
	Estrous cycle		Postpartum period		
	Follicular phase	Luteal phase	15-day PP	30-day PP	45-day PP
<b>Climate condition (s)</b>					
Mild	76.42 $\pm$ 8.05	24.17 $\pm$ 2.14	29.27 $\pm$ 1.8	38.73 $\pm$ 3.30	34.84 $\pm$ 2.3
Hot	78.15 $\pm$ 2.24	28.22 $\pm$ 0.77	64.32 $\pm$ 2.5	47.04 $\pm$ 2.30	41.32 $\pm$ 3.7
<i>P</i> - value	0.75	0.2	0.0001	0.005	0.029
<b>Treatments</b>					
Control	57.00 <sup>B</sup> $\pm$ 2.90	20.74 <sup>B</sup> $\pm$ 1.44	48.32 <sup>B</sup> $\pm$ 5.7	35.45 <sup>B</sup> $\pm$ 2.9	40.08 <sup>B</sup> $\pm$ 2.8
Cr	66.29 <sup>B</sup> $\pm$ 3.45	24.45 <sup>B</sup> $\pm$ 1.56	36.43 <sup>C</sup> $\pm$ 3.2	34.90 <sup>B</sup> $\pm$ 3.3	20.96 <sup>C</sup> $\pm$ 1.5
Se-E	108.56 <sup>A</sup> $\pm$ 8.03	33.39 <sup>A</sup> $\pm$ 2.04	55.65 <sup>A</sup> $\pm$ 3.4	58.23 <sup>A</sup> $\pm$ 1.9	53.19 <sup>A</sup> $\pm$ 3.4
<i>P</i> - value	0.0001	0.0001	0.0001	0.0001	0.0001
<b>Climate x Treatments interactions</b>					
<b>Mild</b>					
Control	41.23 <sup>c</sup> $\pm$ 2.9	15.43 <sup>d</sup> $\pm$ 1.28	21.49 <sup>c</sup> $\pm$ 1.3	25.49 <sup>d</sup> $\pm$ 2.5	30.26 <sup>b</sup> $\pm$ 0.23
Cr	53.80 <sup>c</sup> $\pm$ 4.02	17.38 <sup>d</sup> $\pm$ 0.24	26.04 <sup>c</sup> $\pm$ 3.3	29.43 <sup>d</sup> $\pm$ 3.7	21.05 <sup>b</sup> $\pm$ 1.12
Se-E	134.23 <sup>a</sup> $\pm$ 11.47	39.72 <sup>a</sup> $\pm$ 2.98	40.27 <sup>b</sup> $\pm$ 1.4	61.27 <sup>a</sup> $\pm$ 3.6	53.22 <sup>a</sup> $\pm$ 1.1
<b>Hot</b>					
Control	72.78 <sup>b</sup> $\pm$ 4.47	26.06 <sup>c</sup> $\pm$ 1.35	75.14 <sup>a</sup> $\pm$ 2.2	45.41 <sup>bc</sup> $\pm$ 3.5	49.91 <sup>a</sup> $\pm$ 0.41
Cr	78.77 <sup>b</sup> $\pm$ 2.27	31.52 <sup>b</sup> $\pm$ 1.01	46.81 <sup>b</sup> $\pm$ 3.4	40.38 <sup>c</sup> $\pm$ 5.1	20.88 <sup>b</sup> $\pm$ 2.8
Se-E	82.89 <sup>b</sup> $\pm$ 4.28	27.07 <sup>c</sup> $\pm$ 1.11	71.02 <sup>a</sup> $\pm$ 1.6	55.32 <sup>ab</sup> $\pm$ 1.1	53.16 <sup>a</sup> $\pm$ 6.9
<i>P</i> - value	0.0001	0.0001	0.0001	0.002	0.008

Means with different letters (A, B and C or a, b, c,) in the same column are significantly different at (P<0.05).

On the other hand, treating does with Cr throughout postpartum period did not reveal any enhancement in E<sub>2</sub>-17 $\beta$  levels neither in hot nor mild climates. However, Se-E injection showed significant enhancement in E<sub>2</sub>-17 $\beta$  during days of postpartum with the highest mean value of 58.23pg/ml on day-15 postpartum. Inspection data of climate and treatment interactions, the increase of E<sub>2</sub>-17 $\beta$  due to Se-E on 15<sup>th</sup>, 30<sup>th</sup> and 45<sup>th</sup> days of PP was observed only under mild climate conditions (Table 3).

## DISCUSSION

### *Effect of climate conditions and treatments on progesterone levels.*

Dietary use of a chromium supplementation for approximately 2 months in peak and early lactation cows under natural high ambient temperatures had no effect on progesterone levels (Nikkhah *et al.* (2011). In this respect, the present Cr results of estrous cycle and pregnancy under hot conditions are in agreement with those of Nikkhah *et al.* (2011) but not with results of postpartum since it was almost lower in progesterone than control. The obtained postpartum results agreed with the findings of Yang *et al.* (1996) who found that progesterone levels did not differ in Cr-supplemented cows from control postpartum cows at week 2, while progesterone level at postpartum week 6 was significantly lower in treated cows than control with value about 2.57 vs. 3.34 ng/ml, respectively. Furthermore, Burton *et al.* (1995) found no significant effect of Cr supplementation on p4 at weeks -1, 0, 1 and 2 post calving, although Cr-supplemented cows tended to have lower P4 concentrations that declined more rapidly post calving than control cows. On the other side, Ganie *et al.* (2014) stated that supplementation of Se (0.2 ppm Se selenite) had no effect (P<0.05) on serum P4 profile in buffalo heifers during estrous cycle compared with control group. The present results agreed with the later authors during estrous cycle at hot and mild season (Table2).

In pregnant Baladi goats observed that Se-E increased serum P4 during early, mid- and late-pregnancy (Esa, 2011 and Habeeb *et al.*,2012). In the same trend, plasma P4 concentration increased during pregnancy, reaching its highest level at 19-20<sup>th</sup> weeks and then declined during the last 2 weeks before parturition (Krajnicakova *et al.*, 2003). In addition, Kamada *et al.* (2014) in pregnant heifers found that Se supplementation (0.3 ppm sodium selenite) increased plasma P4 in the 29–39 weeks of pregnancy from 4.98 to 6.86ng/ml, on average. The authors suggested that Se contributes to maintaining the function of the corpus luteum and/or placenta in the latter period of pregnancy. Postpartum results clearly showed highly significant increase in P4 concentration at 30<sup>th</sup> day postpartum due to Se-E injection especially under hot climate conditions. These are in the same trend of Khan *et al.* (2015) in buffalo. The authors estimated progesterone from 30 days postpartum to evaluate the effect of vitamin E and mineral supplementation on the initiation of cyclicity in the experimental buffaloes, Se-E group showed early initiation of cyclicity (32 days postpartum) compared with control group (35 days postpartum). Cyclicity in most of the animals might have been initiated earlier than 30 days as was evident from P4 concentration. Mavi *et al.* (2006) found that supplementation of vitamin E and selenium to Murrah buffaloes during pre-partum period resulted in early initiation of postpartum ovarian activity and early exhibition of first postpartum heat, as well as, duration of first P4 rise of over 1 ng/ ml of plasma was found to be significantly longer in Se-E supplemented buffaloes.

### *Effect of climate conditions and treatments on estradiol-17 $\beta$ levels.*

Feed supplementation with 0.25, 0.5 and 1.0 mg Cr kg<sup>-1</sup> dry matter, respectively, significantly developed graffian follicles and estrus incidence of dairy heifers (Biswas *et al.*, 2006). In this respect, the cur-

rent increase of estradiol-17 $\beta$  throughout estrous cycle due to Cr goes in harmony with those of **Biswas et al. (2006)**. This increase in E<sub>2</sub> concentrations during estrus may reflect Cr enhancing effect on graffian follicles growth and maturation and subsequently released amount of estradiol-17 $\beta$  from granulose cells. In addition, it may be due to minimization of the early atresia of medium sized follicles caused by heat stress (**Roth et al.,2000**) and this may attribute the obtained increase of E<sub>2</sub> (P<0.05) at luteal phase under hot season condition.

It is known that heat stress reduces plasma concentrations of E2 and lower follicular E2 concentration, aromatase activity, LH receptor level, and delay ovulation (**Ozawa et al., 2005**). Some effects of heat stress may involve adrenocorticotrophic hormone (ACTH) and increase cortisol secretion. ACTH has been reported to block E2 induced sexual behavior (**Habeeb et al., 2012 and Hein and Allrich, 1992**). Increased corticosteroid secretion has been suggested, because this can inhibit GnRH and thus LH secretion (**Gilad et al., 1993**). Many studies confirmed the decrease of cortisol due to Cr administration, which in turn may act on alleviate the adverse effects of heat stress on reproductive hormones (**Pechova et al.,2002, Louise, 2003 and Soltan et al., 2012**).

Cr exerts a significant influence on follicular maturation and luteinizing hormone. Postpartum results indicated that Cr supplementation had no effect on enhancing estradiol-17 $\beta$  levels during postpartum period neither under mild nor hot season conditions (**Tuormaa, 2000**). These results are in agreement with those of **Burton et al. (1995)** who found no significant effect of Cr supplementation on estradiol on weeks -1, 0, 1 and 2 post calving. On the other side, Se-E increased E<sub>2</sub> concentrations during estrus and postpartum period not only under mild conditions but also under hot season. Administration of 0.5-1.5 mg/ml selenium and 50 mg vitamin E provides a real strong influence to the increased concentrations of estrogen (E<sub>2</sub>) during estrus

in dairy cows compared with control group (9.9 vs. 7.81 pg/ml, respectively) (**Prasadini, et al., 2014**). The supplementation of vitamin E-selenium to repeat breeder and anoestrous buffaloes relieved the oxidative stress as shown by the reduced levels of lipid peroxidation, activities of superoxide dismutase and glucose-6-phosphate dehydrogenase along with increased vitamin E and  $\beta$ -carotene and improved blood biochemical composition in vitamin E-Se supplemented animals (**Nayyar and Jindal, 2010**).

In conclusion, Cr showed slight increment in E2-17 $\beta$  concentrations at estrous but not in postpartum period, however it enhanced (P<0.05) P4 levels during different stages of goats reproductive cycle. Se-E increased (P<0.05) E2 levels at estrous, pregnancy and PP periods especially under mild climate conditions, the same trend was obtained for P4 concentrations, except postpartum period.

## REFERENCES

- **Al-Katanani, Y. M.; Paula-Lopes, F.F. and Hansen, P.J. (2002)**: Effect of season and exposure to heat stress on oocyte competence in Holstein cows. *J. Dairy Sci.*, 85: 390.
- **Biswas, P.; Haldar, S.; Pakhira, M.C.; Ghosh, T.K. and Biswas, C. (2006)**: Efficiency of nutrient utilization and reproductive performance of pre-pubertal anestrous dairy heifers supplemented with inorganic and organic chromium compounds. *J. Sci. Food Agric.*, 86:804.
- **Bonomi, A.; Quarantelli, A.; Bonomi, B. M. and Orlandi, A. (1997)**: The effects of organic chromium on the productive and reproductive efficiency of dairy cattle (in Italy). *Rivista di Scienza dell'Alimentazione*, 26: 21.
- **Buchanan-Smith, J.G.; Nelson, E.C. and Tillman, A.D. (1969)**: Effect of vitamin E and selenium deficiencies on lysosomal and cytoplasmic enzymes in sheep tissues. *J. Nutr.*, 99:387.
- **Burton, J.L.; Nonnecke, B.J.; Elsasser, T.H.; Mallard,**



- B.A.; Yang, W.Z. and Mowat, D.N. (1995):** Immunomodulatory activity of blood serum from chromium-supplemented periparturient dairy cows. *Vet. Immunol. Immunopathol.*, 49(1-2): 29.
- **Chemineau, P.; Baril, G.; Leboeuf, B., Maurel, M.C.; Roy, F.; Pellicer-Rubio, M.; Malpaux, B. and Cognie, Y. (1999):** Implications des progres recents en Physiologie de la reproduction dans l'espece caprine. *INRA Prod Anim.* 12:135.
  - **Duncan, D. B. (1955):** Multiple range and multiple F-test. *Biometrics*, 11: 1.
  - **El-Tarabany, M. S.; El-Tarabany, A.A. and Atta, M. A. (2018):** Effect of season on hormonal profile and some biochemical parameters at different stages of estrous cycles in Baladi goats, *Biological Rhythm Research*, DOI: 10.1080/09291016.2018.1440775.
  - **El-Tarabany, M. S; El-Tarabany, A.A. and Atta, M.A. (2017):** Physiological and lactation responses of Egyptian dairy Baladi goats to natural thermal stress under subtropical environmental conditions. *Int. J. Biometeorol.*, 61: 61.
  - **Elvinger, F.; Natzke, R.P. and Hansen, P.J. (1992):** Interactions of heat stress and bovine somatotropin affecting physiology and immunology of lactating cows. *J. Dairy Sci.*, 75: 449.
  - **Emesih, G.C.; Newton, G.R. and Weise, D.W. (1995):** Effect of heat stress and oxytocin on plasma concentration of progesterone and 13, 14 dihydro-15-keptoprostaglandin F<sub>2</sub> in goats. *Small Ruminant Research*, 16: 133.
  - **Esa, R.A. (2011):** Effect of vitamin E and selenium supplementation on reproductive efficiency and immune response of Baladi goats under Southern Sinai. Ph.D., thesis, Women's Collage for Arts, Science and Education, Ain Shams Univ., Cairo, Egypt.
  - **Fatet, A.; Maria-Teresa, P. and Leboeu, B. (2011):** Reproductive cycle of goats. *Anim. Repr. Sci.*, 124: 211.
  - **Ganie, A.A.; Baghel, R.P.; Mudgal, V.; Aarif, O. and Sheikh, G.G. (2014):** Effect of selenium supplementation on reproductive performance and hormonal profile in buffalo heifers. *Indian J. Anim. Res.*, 48 (1): 27.
  - **Gilad, E.; Meidan, R.; Berman, A.; Graber, Y. and Wolfenson, D. (1993):** Effect of heat stress on tonic and GnRH-induced gonadotrophin secretion in relation to concentration of oestradiol in plasma of cyclic cows. *J. Reprod. Fertil.*, 99: 315.
  - **Hansen, J.P. (2009):** Effects of heat stress on mammalian reproduction. *Phil.Trans. R. Soc. B.*, 364: 334.
  - **Hein, K.G. and Allrich, R.D. (1992):** Influence of exogenous adrenocorticotropic hormone on estrous behavior in cattle. *J. Anim. Sci.*, 70:243.
  - **Jordan, E.R. (2003):** Effects of heat stress on reproduction. *J. Dairy Sci.*, 86: (Suppl. E): 104.
  - **Habeeb, A.A.M; Teama, F.E.I. and EL-Tarabany, A.A. (2012):** Effect of adding selenium and vitamin E to the diet on reproductive traits of female zaraibi goats and growth of their kids. *Isotope. & Rad. Res.*, 44(3): 693.
  - **Kamada, H.; Nonaka, I.; Takenouchi, N. and Amari, M. (2014):** Effects of selenium supplementation on plasma progesterone concentrations in pregnant heifers. *Anim. Sci. J.*, 85(3): 241.
  - **Khan, H.M.; Mohanty, T.K.; Bhakat, M.; Gupta, A.K.; Tyagi, A.K. and Mondal, G. (2015):** Effect of vitamin-E and mineral supplementation on biochemical profile and reproductive performance of buffaloes. *Buffalo Bulletin*, 34(1):63.
  - **Krajnicakova, M.; Kovac, G.; Kostecky, M.; Valocky, I.; Maracek, I.; Sutiakova, I. and Lenhardt, L. (2003):** Selected clinical- biochemical parameters in the puerperal period of goats. *Bulletin of Veterinary Institute Pulawy*, 47: 177.
  - **Louise, N., (2003):** Postpartum changes in hormones and metabolites during early lactation in summer and winter calving Holstein cows. MS Thesis, Department

of Animal Science, Graduate Faculty of North Carolina State University, Raleigh, NC, USA.

- **Marai, I.F.M.; Bahgat, L.B.; Shalaby, T.H. and Abdel-Hafez, M.A. (2000):** Fattening performance, some behavioral traits and physiological reactions of male lambs fed concentrates mixture alone with or without natural clay, under hot summer of Egypt. *Ann. Arid Zone*, 39 (4): 449.
- **Mavi, P. S.; Pangaonkar, G. R. and Sharma, R. K. (2006):** Effect of vitamin E and selenium on postpartum reproductive performance of buffaloes. *Indian J. Anim. Sci.*, 76(4): 308.
- **Nayyar, S. and Jindal, R. (2010):** Essentiality of antioxidant vitamins for ruminants in relation to stress and reproduction. *Iran. J. Vet. Res.*, 11(1): 1.
- **Nikkhah, A.; Mirzaei, M.; Khorvash, M.; Rahmani, H. R. and Ghorbani, G. R. (2011):** Chromium improves production and alters metabolism of early lactation cows in summer. *J. Anim. Physiol. Ann.*, 95:81.
- **NRC, (2007):** Nutrient Requirements of Small Ruminants: Sheep, Goats, Cervids, and New World Camelids. Washington, DC, Natl. Acad. Press.
- **Ozawa, M.; Tabayashi, D.; Latief, T. A.; Shimizu, T.; Oshima, I. and Kanai, Y. (2005):** Alterations in follicular dynamics and steroidogenic abilities induced by heat stress during follicular recruitment in goats. *Reproduction*, 129: 621.
- **Pechova, A. and Pavlata, L. (2007):** Chromium as an essential nutrient: a review. *Veterinari Med.*, 52(1): 1.
- **Pechova, A.; Cech S.; Pavlata L. and Podhorsky A. (2003):** The influence of chromium supplementation on metabolism, performance and reproduction of dairy cows in a herd with increased occurrence of ketosis. *Czech J. Anim. Sci.*, 48: 348.
- **Pechova, A.; Pavlata, L. and Illek, J. (2002):** Metabolic effects of chromium administration to dairy cows in the period of stress. *Czech J. of Anim. Sci.*, 47: 1.
- **Prasadini, W. A; Rahayu, S. and Djati, M. S. (2014):** Level of estrogen and cervical mucus pH as indicator of estrus after calving towards the provision of selenium vitamin E on dairy cow Frisien Holstein (FH). *Int. J. Chem. Tech. Res.*, 7(1): 190.
- **Roth, Z.; Meidan, R.; Braw-Tal, R. and Wolfenson, D. (2000):** Immediate and delayed effects of heat stress on follicular development and its association with plasma FSH and inhibin concentration in cows. *J. Reprod. Fertil.*, 120: 83.
- **Sartori, R.; Sartor-Bergfelt, R.; Mertens, S.A.; Guenther, J.N.; Parrish, J.J. and Wiltbank, M.C. (2002):** Fertilization and early embryonic development in heifers and lactating cows in summer and lactating and dry cows in winter. *J. Dairy Sci.*, 85: 2803.
- **SAS (1998):** Statistical analysis system user's guide, Release 6.0 ed. 4<sup>th</sup> ed, SAS Institute Inc. Cary, NC, USA.
- **Soltan, M. A.; Almujaali, A.M.; Mandour, M.A. and El-Shinway, A.M. (2012):** Effect of dietary chromium supplementation on growth performance, rumen fermentation characteristics and some blood serum units of fattening dairy calves under heat stress. *Pakistan J. Nutr.*, 11(9): 751.
- **Surai, P.F. (2006):** Selenium in Nutrition and Health. 1<sup>st</sup> ed. Nottingham: Nottingham Univ. Press.
- **Tuormaa, T. E. (2000):** Chromium selenium copper and other trace minerals in health and reproduction. *J. Orthomol. Med.*, 15: 145.
- **Williams, J. E.; Myers, J.L.; Richard, C.R. and Grebing, S.E. (1994):** Influence of yeast culture, chromium, and thermal challenge on N and mineral balance in lambs. *J. Anim. Sci.*, 72(Suppl. 2): 86.
- **Wise, M.E.; Armstrong, D.V.; Huber, J.T.; Hunter, R. and Wiersma, F. (1988):** Hormonal alterations in the lactating dairy cow in response to thermal stress. *J. Dairy Sci.*,

71: 2480.

- **Yang, W.Z.; Mowat, D.N.; Subiyatno, A. and Liptrap, R.M. (1996):** Effects of chromium supplementation on early lactation performance of Holstein cows. *Can. J. Anim. sci.*, 6: 221.

## تأثير الكروميوم والسيلينيوم- ه علي مستويات البروجستيرون والإستراديول-17بيتا خلال الدورة التناسلية لإناث الماعز البلدي تحت الظروف المصرية

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إجريت هذه التجربة لدراسة تأثير معاملة اناث الماعز المحلية بالكروميوم والسيلينيوم-ه علي مستويات هرموني البروجستيرون والإستراديول 17-بيتا خلال دورة الشبق ومرحلة الحمل وما بعد الولادة للحيوانات كطرق لتحسين أداء الماعز المصرية تحت الظروف المحلية .  
وقد استخدم في هذا البحث 72 عنزة بلدي ناضجة بمتوسط عمر 2-3 سنوات ومتوسط وزن الجسم 25 كجم خلال موسم دراسه وهما ظروف المناخ المعتدل والحر بحيث 36 حيوان في كل موسم وفي كل موسم قسمت الحيوانات عشوائيا الي ثلاث مجاميع الاولي بدون معاملات (مجموعه ضابطه) والمجموعه الثانيه تم معاملتها بمادة الكروميوم - في صورة كلوريد الكروميوم- وبمعدل 0.8 ملجم/رأس/ يوم (مجموعه الكروميوم) أما المجموعه الثالثه تم حقنها في العضل بـ 2مل مرتين اسبوعياً من مخلوط السيلينيوم+فيتامين ه (فايتسولين) وهذه الكمية تعادل 0.5 ملجم من السيلينيوم و 10.7 وحدة دولية فيتامين ه /رأس/يوم.  
وكانت الحيوانات موضوعة في حظائر نصف مفتوحة خلال فترة التجربة وسمح لها بالرعي لمدة خمسة ساعات يومياً علي الأقل وقد تم تقدير مستوي الهرمونات في سيرم الدم خلال المراحل التناسليه المختلفه.  
وقد أوضحت النتائج أن مايلي:

1. ارتفاع معنوي في مستوي هرمون الإستراديول خلال مرحلتي النمو الحويصلي والجسم الأصفر وفترة ما بعد الولادة نتيجة المعاملة بالسيلينيوم-ه مقارنة بالكنترول،
2. كانت مستويات الهرمون اعلي تحت ظروف الجو المعتدل عن الموسم الحر داخل نفس المجموعه خلال معظم المراحل التناسليه المختلفه.
3. المعاملة بالكروميوم ادت الي ارتفاع -وان كان غير معنوي- في مستوي الإستراديول خلال مرحلتي دورة الشبق ولم يكن للكروميوم تاثير واضح علي مستوي الهرمون خلال فترة ما بعد الولادة لاناث الماعز المحلية تحت ظروف المناخ المعتدل او الحر.
4. أظهرت المعاملة بالكروميوم تأثيراً معنوياً علي مستوي هرمون البروجستيرون خلال دورة الشبق والحمل مقارنة بالكنترول في حين لم يكن له تأثير واضح خلال فترة ما بعد الولادة بخلاف اليوم الـ 15 ما بعد الولادة تحت ظروف المناخ المعتدل.
5. المعاملة بالسيلينيوم-ه ادت الي زيادة تركيز البروجستين بشكل معنوي مقارنة بالكنترول خلال مرحلة الجسم الأصفر وفترة الحمل وذلك في الجو المعتدل ولم يكن له تأثير خلال فترة ما بعد الولادة عدا اليوم الـ 30 تحت ظروف الجو الحر.

1. قسم الإنتاج الحيواني -كلية الزراعة- جامعة بنها.

2. قسم التطبيقات البيولوجية - شعبة تطبيقات النظائر المشعة - مركز البحوث النووية - هيئة الطاقة الذرية.