Isolation and Antibiogram of *Escherichia coli* from Canine Pyometra in Puducherry Region

Manivasagam Vignesh¹, Jayalakshmi Vasu²*, Devanathan Nivedha², Mouttou Vivek Srinivas², Shalini Iyyanar³, Hirak Kumar Mukhopadhyay²

ABSTRACT

Pyometra is a life-threatening and hormone induced cystic endometrial hyperplasia seen predominantly in older bitches, complicated with secondary bacterial infections. This study was performed with the objective of isolation and antibiogram of *Escherichia coli* from canine pyometra. Vaginal swabs were collected aseptically from 20 canine pyometra cases, presented to Veterinary Clinical Complex, Puducherry (India) for identification of *E. coli* isolates. Based on colony characters, microscopic observation, biochemical tests, 12 (60%) *E. coli* isolates were identified. All the 12 phenotypically positive *E. coli* isolates were confirmed further by PCR using the primer targeting *alr* gene. *E. coli* isolates were found sensitive to Gentamicin (100%), Ceftriaxone (83%), Enrofloxacin (75%) and Ciprofloxacin (66%), but were resistant to commonly used antibiotics like Amoxyclav (100%), Ceftazidime (58%) and Cefotaxime (42%). The results provided an authentic and reliable information to choose an effective antibiotic to treat *E. coli* infection causing canine pyometra.

Keywords: Antibiogram, Canine, E. coli, Hormonal imbalance, Pyometra,

Ind J Vet Sci and Biotech (2022): 10.48165/ijvsbt.18.5.27

INTRODUCTION

anine pyometra is a life-threatening condition seen predominantly in middle-aged to older aged bitches during diestrual phase. It is characterized by the accumulation of pus within the uterine lumen, because of progesterone dominance. This hormonal imbalance occurs during the luteal phase of estrous cycle when the uterine immunity is low and the contaminating microorganisms dominate over the protective mechanisms of the female reproductive tract (Roberts, 1986; Johnston et al., 2001; Niyas et al., 2020). Normally dog uterus does not accomodate any microbes but they migrate during the proestrous and estrous phases through vagina and cervix (Watts et al., 1996; Niyas et al., 2020). High or prolonged ovarian production of progesterone causes cystic endometrial hyperplasia and inhibition of myometrial contractility. This abnormal uterine environment allows bacterial colonization to cause pyometra. The most common bacterial infection of canine pyometra is reported to be Escherichia coli.

E. coli is a normal inhabitant of the intestinal tract of humans and animals, and considered to be one of the notorious multi-drug resistant bacterial pathogens causing variety of clinical condition in different animal species (Sangeetha *et al.*, 2016; Tamilarasu *et al.*, 2020). *E. coli* has high affinity to the epithelium and endometrium of the urinary tract especially when sensitized by a high level of progesterone and leads to canine pyometra (So *et al.*, 2000; Chen *et al.*, 2003; Niyas *et al.*, 2020). It is important to identify these *E.coli* causing pyometra cases, so as to proceed for the early identification, diagnosis and appropriate antibiotic intervention to avoid fatal consequences in animals. Therefore, this study was aimed to isolate and identify *E. coli* from canine pyometra cases in Puducherry and to determine their antibiogram ¹Department of Veterinary Clinical Complex, Rajiv Gandhi Institute of Veterinary Education and Research, Kurumbapet, Puducherry-605 009, India.

²Department of Veterinary Microbiology, Rajiv Gandhi Institute of Veterinary Education and Research, Kurumbapet, Puducherry– 605 009, India.

³Department of Veterinary Gynaecology and Obstetrics, Rajiv Gandhi Institute of Veterinary Education and Research, Kurumbapet, Puducherry– 605 009, India.

Corresponding Author: Jayalakshmi Vasu, Department of Veterinary Microbiology, Rajiv Gandhi Institute of Veterinary Education and Research, Kurumbapet, Puducherry– 605 009, India, Phone: 8754782146, e-mail: drjayalakshmivasu@gmail.com

How to cite this article: Vignesh, M., Vasu, J., Nivedha, D., Srinivas, M. V., Iyyanar, S., & Mukhopadhyay, H. K. (2022). Isolation and Antibiogram of Escherichia coli from Canine Pyometra in Puducherry Region. Ind J Vet Sci and Biotech. 18(5), 130-133.

Source of support: None

Conflict of interest: None

Submitted: 18/08/2022 Accepted: 05/11/2022 Published: 10/11/2022

profile. The long treated and recurrent cases were subjected to ABST and accordingly the therapy followed.

MATERIALS AND METHODS

Collection and Processing of Vaginal Swab Samples

A total of 20 clinical cases of canine pyometra, presented to Department of Veterinary Clinical Complex, Rajiv Gandhi Institute of Veterinary Education and Research (RIVER), Puducherry, India were used for the study. The condition was diagnosed based on history, clinical signs and ultrasonography. Sterile cotton swabs individually packed in

© The Author(s). 2022 Open Access This work is licensed under a Creative Commons Attribution-Non Commercial-No Derivatives 4.0 International License.

polypropylene tube (M/s Hi-Media, Mumbai) were used for collection of vaginal swabs from dogs. The samples, after collection, were transported to Department of Veterinary Microbiology, of the Institute within 2 h and immediately processed for culture and isolation.

Isolation and Identification of E. coli

Each vaginal swab from the affected bitches was subjected to enrichment in 5 mL of Luria broth individually in sterile test tube, for 18 h at 37°C. A loopful of the enriched culture was streaked onto MacConkey's agar and incubated at 37°C for 24 h. The pink coloured colonies (Lactose fermenter, Fig. 1a) obtained were subjected to Gram's staining. The Gram negative bacilli were taken up for further study following standard method in practice.

The lactose fermenting colonies were sub-cultured onto Eosin Methylene Blue (EMB) agar, in which metallic green sheen colonies (Fig. 1b) were produced by *E. coli* (Kavitha *et al.*, 2013). The conventional biochemical tests such as Catalase, Oxidase, Indole, Methyl red, Voges-proskauer, Citrate, Urease and Triple sugar iron (TSI) agar tests were used for the phenotypic identification of *E. coli* as described by Krieg and Holt (1984). The phenotypically positive *E. coli* isolates were further confirmed by polymerase chain reaction (PCR).

Polymerase Chain Reaction for Detection of E. coli

The preparation of template DNA from *E. coli* strains was carried out as described by Zhang *et al.* (2015). The PCR was carried out with primers targeting *alr* genes specific for *E.*



Fig. 1: (a) Lactose fermenting colonies in MacConkey's agar, (b) Metallic green sheen colonies in EMB agar.

coli with the product size of 366 bp as per Yokoigawa *et al.* (1999). The PCR amplification was carried out in an automated thermal cycler (Eppendorf Mastercycler, Germany) according to the following protocol: Initial denaturation at 95°C for 6 min followed by 36 cycles of denaturation at 95°C for 20 s, annealing at 72°C for 45 s and extension at 72°C for 45 s and final extension at 72°C for 5 min. The PCR products were analyzed on 1.5% agarose gel electrophoresis.

Antibiotic Sensitivity Test (ABST) of *E. coli* Isolates

Isolated *E. coli* organisms were cultured on Mueller Hinton agar to obtain a smooth, homogeneous lawn culture and subjected to antibiotic sensitivity testing using 8 different antibiotics by the disc diffusion method (Bauer *et al.*, 1966). The antimicrobial agents used for the sensitivity testing were Gentamicin (GEN, 10 μ g), Amoxyclav (AMC, 30 μ g), Ciprofloxacin (CIP, 5 μ g), Enrofloxacin (EX, 5 μ g), Cefotaxime (CTX, 30 μ g), Ceftazidime (CAZ, 30 μ g), Doxycycline (DO, 10 μ g) and Ceftriaxone (CTR, 30 μ g). The interpretation of zone diameter was carried out according to Clinical Laboratory Standard Institute (CLSI, 2019).

RESULTS AND **D**ISCUSSION

Based on colony characters, microscopic observation, biochemical tests, 12/20 (60%) *E. coli* isolates were identified. All the 12 (100%) phenotypically detected *E. coli* isolates in the study were further confirmed by PCR with *E. coli* species specific primers targeting *alr* gene (Fig. 2). Similarly Chen *et al.* (2003) and Xavier *et al.* (2022) by using both phenotypic and genotypic method detected 100% (24/24) and 56% (40/72) of *E. coli* isolates from pyometra cases in bitches.



Fig. 2: Agarose gel electrophoresis showing the results of PCR amplified product of *alr* gene of *E. coli* with size 366 bp. Lane 1 & 2 Negative and Positive control, respectively; Lane 3, 4, 5 and 6 *E. coli* isolates; Lane 7 100 bp ladder.

131

Isolation and Antibiogram o	of Escherichia coli from	Canine Pyometra
-----------------------------	--------------------------	-----------------

Table 1: Antibiogram of the E. coli isolates (n=12)				
Antibiotic	Sensitive	Intermediate	Resistant	
Ceftriaxone	83% (10)	-	17% (2)	
Ciprofloxacin	67% (8)	8% (1)	25% (3)	
Enrofloxacin	75% (9)	-	25% (3)	
Amoxyclav	-	-	100 % (12)	
Doxycycline	8% (1)	67% (8)	25% (3)	
Ceftazidime	25% (3)	17% (2)	58% (7)	
Gentamicin	100% (12)	-	-	
Cefotaxime	33% (4)	25% (3)	42% (5)	

a. 1 - 2

*(n)- no. of isolates

Antibiogram pattern of the E. coli isolates obtained from pyometra in this study showed resistant to most commonly used antibiotics (Table 1, Fig. 3). All the E. coli isolates were resistant against Amoxyclav (100%) followed by Ceftazidime (58%) and Cefotaxime (42%). Most sensitive antibiotics were found to be Gentamicin (100%), Ceftriaxone (83%), Enrofloxacin (75%) and Ciprofloxacin (66%). Gentamicin was detected as the most effective antibiotic which was in agreement with the previous reports of Bassessar et al. (2013) and Agostinho et al. (2014). But Gentamicin, being moderately nephrotoxic restricts its clinical usage in canine. Second most effective antibiotic was Ceftriaxone followed by Quinolones can be included for antibiotic therapy. Similar result was found by Rocha et al. (2021), who reported that the E. coli isolates were found sensitive to Ceftrioxone (83.33%) and Quinolones (66.67%), while Bassessar et al. (2013) reported about 55% to 65% sensitivity to Quinolone antibiotics.



Fig 3: Antibiotic sensitivity Test (ABST) result of E. coli isolates

CONCLUSION

The present study showed that E. coli is the most common bacteria isolated from pyometra cases in different breeds of bitches in Puducherry region. Gentamicin and Ceftrioxone were detected as the most effective antibiotics and the isolates showed resistant to commonly used antibiotics such as Ceftazidime and Cefotaxime. Further studies are needed to determine the virulence genes of the bacteria isolated from pyometra cases in order to assess their pathogenic potential.

ACKNOWLEDGEMENTS

The authors are thankful to the Dean, Rajiv Gandhi Institute of Veterinary Education and Research, Puducherry, India for providing necessary funds and facilities to carry out this study.

REFERENCES

- Agostinho, J.M., de Souza, A., Schocken-Iturrino, R.P., Beraldo, L.G., Borges, C.A., Avila, F.A., & Marin, J.M. (2014). Escherichia coli strains isolated from the uteri horn, mouth, and rectum of bitches suffering from pyometra: Virulence factors, antimicrobial susceptibilities, and clonal relationships among strains. International Journal of Microbiology, 2014, 979584. http://dx.doi.org/10.1155/2014/979584.
- Bassessar, V., Verma, Y., & Swamy, M. (2013). Antibiogram of bacterial species isolated from canine pyometra. Veterinary World, 6, 546-549.
- Bauer, A. W., Kirby, W. M., Sherris, J. C., & Turck, M. (1966). Antibiotic susceptibility testing by a standardized single disk method. American Journal of Clinical Pathology. 45(4), 493-496.
- Krieg, N.R. & Holt, J.G., (1984). Bergey's Manual of Systematic Bacteriology. 1st ed., Vol. 1, Williams and Willkins, Baltimore. London.
- Chen, Y.M., Wright, P.J., Lee, C., & Browning, G.F. (2003). Uropathogenic virulence factors in isolates of Escherichia coli from clinical cases of canine pyometra and feces of healthy bitches. Veterinary Microbiology, 94(1), 57-69.
- Clinical and Laboratory Standards institute (CLSI, 2019) Performance Standards for Antimicrobial Susceptibility Testing: 24th Informational Supplement. M100-S24. Clinical and Laboratory Standards Institute, Wayne, PA, USA.
- Johnston, S.D., Kustritz, M.V., & Pn, O. (2001). Canine and Feline Theriogenology. 1st edn. W.B. Saunders, Philadelphia, pp. 207-220.
- Kavitha, J.R. and Devasena, T. (2013). Molecular and bacteriological examination of cow milk in coliform mastitis. IOSR Journal of Pharmacy and Biological Sciences, 6(2), 34-40.
- Niyas, E., Reshma, S., Shravya, G.S., Jayakumar, C.L., Kumar, R.H., Sarika, N., & Easaw, A.M. (2020). Antibiogram of Isolated Organisms in Canine Pyometra. International Journal of Current Microbiology and Applied Sciences, 9, 263-268.
- Roberts, S.J. (1986). Veterinary Obstetrics and Genital Disease. 3rd edn. Published by the Author, Woodstock, VT. Ithaca, New York, pp. 734-739.
- Rocha, R.A., Ribeiro, W.M., Almeida, J.A. de, Santos, A.L., Fernandes, M.R., Barbosa, M.S., Moraes Filho, A.V. de, Carneiro, L.C., & Silva,



C.A. da. (2021). Detection of resistance genes in pyometra isolated bacteria in bitches. *The Brazilian Journal of Veterinary Research and Animal Science, 58*, e173908.

- Sangeetha, B., Antony, P.X., Mukhopadhyay, H.K., Madhusoodanan Pillai, R, Thanislass, J. Vijayalakshmi, P. & Mouttou V.S. (2016). Genetic characterization of Fluroquinolone Resistant Escherichia coli associated with bovine mastitis in India. *Veterinary World*, *9*, 705-709.
- So, H.L., JongKi, C., nari, S., Hyesoo, K., HwanYul, Y., HanSang, Y., Kang-nam, L., ByeongChun, L., & WooSuk, H. (2000). Identification and antimicrobial susceptibility of bacteria from the uterus of bitches with pyometra. *Korean Journal of Veterinary Research*, 40(4), 763-767
- Tamilarasu, S., Jayalakshmi, V., Vivek Srinivas, V.M., Vinodh Kumar, O.R., Antony, P.X., & Mukhopadhyay, H,K. (2020). Antibiotic susceptibility pattern of extended spectrum β-lactamase (ESBL) producing *Escherichia coli* isolated from dogs. *Veterinary Research International*, 8(2), 58-61.

- Watts, J.R., Wright, P.J., & Whithear, K.C. (1996). Uterine, cervical and vaginal microflora of the normal bitch throughout the reproductive cycle. *The Journal of Small Animal Practice*, *37*(2), 54-60.
- Xavier, R.G.C., da Silva, P.H.S., Trindade, H.D., Carvalho, G.M., Nicolino, R.R., Freitas, P.M.C., & Silva, R.O.S. (2022). Characterization of *Escherichia coli* in dogs with pyometra and the influence of diet on the intestinal colonization of extraintestinal pathogenic *E. coli* (ExPEC). *Veterinary Sciences*, 9, 245.
- Yokoigawa, K., Inoue, K., Okubo, Y. & Kawai, H. (1999). Primers for amplifying an alanine racemase gene fragment to detect *E. coli* strains in foods. *Journal of Food Science*, *64*, 571-575.
- Zhang, H., Zhou, Y., Guo, S., & Chang, W. (2015). High prevalence and risk factors of fecal carriage of CTX-M type extendedspectrum beta-lactamase-producing Enterobacteriaceae from healthy rural residents of Taiwan, China. *Frontiers in Microbiology*, *6*, 239.