Effect of post AI intrauterine treatment with *E. coli* lipopolysachharide and autologous serum on characteristics of cervico-vaginal mucous in subclinical endometritic cows

P. SINGLA¹, JAGIR SINGH², G. S. DHALIWAL^{3†} AND AJEET KUMAR⁴

Department of Animal Reproduction, Gynaecology & Obstetrics Punjab Agricultural University, Ludhiana-141004 (Punjab)

> Received: March 8, 2002 Accepted: September 22, 2003

ABSTRACT

Sub-clinical bacterial endometritis is a common cause of repeat-breeding in cows. The present study was conducted on 36 cows with confirmed sub-clinical endometritis. Based on the treatment protocol, these cows were divided randomly into three groups of 12 animals each. Group 1 (control), Group 2 and 3 cows were infused intrauterine with 20 ml of phosphate buffer saline (PBS), E.coli lipopolysaccharide (LPS) 100µg and E. coli LPS 100µg + 5 ml of autologous serum, respectively, 8-12 hr after insemination. Cervico-vaginal mucus (CVM) was collected before artificial insemination (AI) and 12 hr after treatment. It was observed that colour and consistency of CVM became normal in significant number of animals. A significant decrease in pH of CVM was also observed in treated animals.

Key Words: Lipopolysachharide, autologous serum, sub-clinical endometritis, cervico-vaginal mucus, cows

Bacterial endometritis is one of the common causes of altering the physical properties of the CVM (Dohman et al., 1996). Normal CVM is important for sperm transport in genital tract (Rowson et al., 1972). Intrauterine antibiotics/antiseptics have been used for the treatment of endometritis with variable success (Shukla and Pandit, 1989). The disadvantages of antibiotics/antiseptics are compulsory milk disposal, development of microbial resistance, higher cost and inhibition of phagocytic activity of polymorphonuclear cells (Vandeplassche, 1984). Taking these problems in to consideration, alternative therapies have been used for the treatment of uterine infection. One is them is the use of immunomodulators. Oyster glycogen has been used endometritis, but, there was loss of one complete estrous cycle (Anderson et al., 1984). Therefore, LPS was tried as post AI therapy in the sub-clinical endometritis cows in the present study to save one estrous cycle.

Thirty six normal cycling crossbred cows maintained at dairy farm, PAU, Ludhiana; Habowal Dairy Complex and animals brought to PAU Clinics were included in the study.

Sub-clinical endometritis was confirmed on the basis of bacterial culture and isolation from uterine swabs. The animals were divided randomly into three groups of 12 animals each. Group 1 (control), group 2 and group 3 cows were infused intrauterine with 20 ml phosphate buffer saline (PBS), 100 µg of LPS* and 100µg LPS+5ml autologous serum, respectively, 8-10 hrs after insemination. Cervico-vaginal mucus was collected using AI glass pipette and syringe aseptically before and 12 hrs after treatment. Color, consistency and pH of CVM were assessed. Colour was assessed with the naked eye. Consistency was judged and graded as thin (++) and thick (+++). pH was noted with the help of pH paper.

Bacteria isolated from uterine swabs are presented in Table-1. Microorganisms isolated from uterine swabs of cows with subclinical endometritis were mainly E. coli, Staph. aureus and Proteus spp., whereas the incidence of Streptococci spp., Klebsiella spp., Pseudomonas spp., Corynebacterium spp. and Bacillus spp. were relatively less. Almost similar bacterial isolates in bovines with endometritis had been reported by Shukla (1988), Singla et al. (1991) and Singh (1996). On the contrary, other workers reported that Staph. aureus, Actinomyces pyogenes, Streptococcus spp. and Bacillus spp. predominanted in repeat breeding bovines (Sharda et al., 1991, Goswami et al., 1992 and Saini, 1993). More incidences of E. coli and Staph. aureus in the subclinical endometritis could

of

¹Veterinary Officer ²Gynaecologist ³Professor & Head ⁴Assistant Professor.

¹Corresponding author

Table 1. Bacteria isolated from uterine swabs in cows with sub clinical endometritis

Bacterial isolate	No. of isolates	Per cent
E. coli	14	31.11
Staph. aureus	12	26.67
Proteus spp.	8	17.77
Streptococci spp.	4	8.88
Klebsiella spp.	3	6.67
Pseudomonas spp.	2	4.44
Corynebacterium spp.	1	2.22
Bacillus spp.	1	2.22
Total no. of isolates	45	100.00

indicate that, these bacteria are more commonly present in the animals excreta like dung etc. Unhygienic conditions at the time of per-rectal examination or at the time of A.I. may lead to the entry of these organisms into uterus causing subclinical endometritis

Colour, consistency and pH of CVM were recorded before and after treatment and the data is presented in Table-2. Before treatment, CVM was turbid in about 83, 75 and 67% animals in groups 1, 2 and 3, respectively. Upon LPS treatment in group 2, turbidity of CVM decreased significantly (P<0.05) from 75 to 25% animals, whereas, treatment in group 3 showed maximum efficacy in clearance of CVM turbidity. However, PBS treatment in control group did not result any change in the colour of CVM. The cervical mucus of healthy animals is usually colourless and transparent (Roberts, 1986 and Arthur et al., 1989). Cloudy, milky and turbid cervical discharge is generally observed due to uterine and cervical infections (Pandey et al., 1983, Shankar et al., 1984, and Saini, 1993).

Consistency of CVM before treatment was thick (+++) in about 42, 33 and 25% animals in group 1, 2 and 3, respectively. After treatment in group 2 and 3, the consistency of CVM changed to normal (thin, ++) in 83% and 92% animals, respectively. However, the changes in the consistency of CVM were non significant in control group after PBS infusion. In the earlier studies, the incidence of thin mucus in repeat breeder cows was either similar (Deo and Roy, 1971) or was lower (62-66%) than in the normal bovines (87-92%; Bishnoi 1975, and Saini 1993). Enkhia and Kohli (1982), however, reported that the CVM in majority of repeat breeding cows (65%) was watery. The difference in the cause and type of inflammation (Runnells et al., 1965) might probably contribute to the variation in the

[able-2. Colour, consistency and pH of cervico-vaginal mucus before and after treatment with PBS, LPS and LPS + autologous serum.

Group		Pre tr	Pre treatment				Post treatment	ient		
	S	Colour	Consistency	tency	Colour		Consistency	ıcy		
	Clean	Turbid	Thin (++)	Thick (+++)	Hd	Clean	Turbid	Thin (++)	Thick (+++)	Hd
1 (n=12)	2 (16.66%)	10 (83.38%)	7 (58.3%)	5 (41.66%)	7.71 ± 0.05 ^a	2 (16.66%)	10 (83.38%)	8 (66.66%)	(33.33%)	7.68 ± 0.06
2 (n=12)	3 (25.00%)	9 (75.00%)	8 (66.66%)	(33.33%)	7.71±0.048	9 (75.00%)	3 (25.00%)	10 (83.33%)	2 (16.66%)	7.23 ± 0.05^{b}
3 (n=12)	(33.33%)	8 (66.66%)	(75.00%)	(25.00%)	7.68±0.05ª	(91.67%)	(8.33%)	(91.61%)	(8.33%)	7.20 ± 0.04 ^C

Doh

Enkh

Gosw

Hafez

consistency of CVM. The CVM consistency primarily depends upon the degree of cross linkage in the epithelial glycoprotein macromolecules of estrual cervical mucus, which in turn brought forth periodic changes in the penetrability of spermatozoa in cervical canal (Hafez, 1993).

pH of CVM before treatment were 7.71±0.05, 7.71±0.04 and 7.68±0.05 in group 1, 2 and 3, respectively. pH decreased marginally to 7.68±0.06 in control group. However, significant decrease in pH of CVM was observed after treatment in groups 2 (7.23±0.05) and 3 (7.20±0.04). The pH of estrual CVM in repeat breeder cows with subclinical endometritis ranged from 7.40 to 8.00 (Akhtar and Singh, 1979; Wani et al., 1982; Rane et al., 1992 and Saini, 1993). The pH of normal CVM is around 7.00-7.20. It was concluded that treatment with LPS or LPS + autologous serum improved colour, consistency and pH of CVM.

REFERENCES

- Akhtar, M.H. and Singh, B.K. (1979). Livability and fertility rate of spermatozoa in bovine cervical mucus under normal and disease conditions. Indian Vet. J., 56: 112-17.
- Anderson, K.L., Hemeida, N.A. and Frank, A. (1984). Effect of oyster glycogen recovered from the bovine uterus. Proc 10th Int Congr on Anim Reprod and A.I., Urbana Champaign, Illinois, USA: 442-44.
- Arthur, G.H., Noakes, D.E. and Pearson, H. (1989). Veterinry Reproduction and Obstetrics. 6th edn, The English Language Book Society and Bailliere Tindall, London.
- Bishnoi, B.L. (1975). Studies on the physical character of bovine cervical mucus during oestrus. M.V.Sc. thesis, Udaipur Univ., Udaipur (Original not seen. Cited by Enkhia and Kohli 1982).
- Deo, S. and Roy, D.J. (1971). Investigations on repeat breeding cows and buffaloes: Studies on physical properties of cervical mucus. Indian Vet J., 48: 479-84.
- Dohmen, M.J.W., Huszenicza, Gy., Fodor, M., Kulcsar, M., Vamos, M., Porkolab, L., Szilagy, N. and Lohuis, J.A.C.M. (1996). Bacteriology and fertility in healthy postpartum cows and cows with acute endometritis. Proc XIX World Buiatrics Congr, Edinburgh, 8-12 July 1: 238-40
- Enkhia, K.L. and Kohli, I.S. (1982). Note on the physical properties of cervico-vaginal mucus during oestrus in normal and repeat breeding Rathi cows. Indian J Anim Sci., 52: 1239-40.
- Goswami I G, Kher H N, Jhala M K and Devashri H J (1992). Quantitative and qualitative studies on the genital bacteria of buffaloes. Indian J Anim Reprod., 13: 180-82.
- Hafez, E.S.E. (1993). Reproduction in Farm Animals. 6th edn, Lea and Febiger, Philadelphia.

- Hatch, R.D., Feenstra, E.S. and Jennings, L.F. (1979). A bacteriological survey of the reproductive tract of infertile cows. J Am. -Vet. Med. Assoc., 114: 131-33.
- Pandey, S.K., Pandit, R.K. and Chaudhary, R.A. (1983). Repeat breeding cows in relation to physical characteristics of cervical mucus, fertility and treatment. Indian Vet J., 60: 946-47.
- Rane, P.M., Sonawane, S.A., Hukeri, V.B., Manitri, A.M. and Narayanhedkar, S.G. (1992). Studies on biochemical attributes of cervical mucus in normal and repeat breeding crossbred cows. National Symposium on Recent Advances in Clinical Reprod in Dairy Cattle and 10th National Convention, ISSAR, TANVASU, Madras. 8-10th April.
- Roberts, S.J. (1986). Veterinary Obstetrics and Genital Diseases. 3rd edn, Woodstock, Vermont.
- Rowson, L.E.A., Lawson, R.A.S., Moor, R.M. and Baker, A.A. (1972).
 Egg transfer in the cow: Synchronization requirements. J
 Reprod Fertil., 28: 427-31.
- Runnells, R.A., Monlux, W.S. and Monlux, A.W. (1965). Principles of Veterinary Pathology. 7th edn, Scientific Book Agency, Calcutta.
- Saini, P.S. (1993). Studies on etiopathology and modified therapy of bovine endometritis. M.V.Sc. thesis, Punjab Agricultural University, Ludhiana, India.
- Shankar, U., Sharma, M.C., Verma, R.P. and Gupta, O.P. (1984). Physicobiochemical studies of cervical mucus in cyclic and repeat breeding crossbred cattle. Indian J Anim Reprod., 4: 42-44.
- Sharda, R., Moghe, M.N. and Tanwani, S.K. (1991). Antibiotic sensitivity pattern of bacteria isolated from repeat breeding animals. Indian Vet J., 68: 197-200.
- Shukla, S.P. (1988). Studies on biochemical changes in the uterine fluid in relation to bacteriology and histopathology of the uterus in repeat breeding cattle. Ph.D. Thesis, Punjab Agricultural University, Ludhiana, India.
- Shukla, S.P. and Pandit, R.K. (1989). Incidence of repeat breeding and its remedial measures in Gir cows and their crosses. Indian Vet. J., 66: 626-30.
- Singh, J. (1996). Studies on uterine defense modulation and its therapeutic importance in repeat breeding cows. Ph.D. thesis, Punjab Agricultural University, Ludhiana, India.
- Singla, V.K., Verma, H.K., Dwivedi, P.N. and Gandotra, V.K. (1991).
 Bacteriological isolates in repeat breeder cows. Indian J. Anim.
 Sci., 61: 181-82.
- Vandeplassche, M. (1984). Stimulation and inhibition of phagocytosis in domestic animals. Proc 10th Int. Congr Anim Reprod and A.I., Urbana Champaign, Illinois, USA 3: 475-77.
- Wani, G.M., Tripathi, S.S. and Saxena, V.S. (1982). Studies on biochemical properties of cervical mucus in normal and repeat breeding Sahiwal cows. Indian J Anim Hlth., 21: 29-31.