DOI: 10.48165/ijar.2021.42.2.3



ISSN 0970-2997 (Print)

The Indian Journal of Animal Reproduction

The official journal of the Indian Society for Study of Animal Reproduction

Year 2021, Volume-42, Issue-2 (Dec)

ACS Publisher www.acspublisher.com

Therapeutic Outcomes Following Uterine Lavage, Levamisole, $PGF_{2\alpha}$ and their Combinations in Subclinical Endometritic Buffaloes

Narendra Singh¹, Bhoopendra Singh², and Rajesh Kumar^{2*}

¹Department of Veterinary Clinical Complex, R.R. College of Veterinary and Animal Science, Deoli, Tonk, RAJUVAS, Rajasthan-304 804, India ²Department of Veterinary Gynaecology & Obstetrics, College of Veterinary Science and AH, ANDUAT, Ayodhya - 224 229 (UP), India

ABSTRACT

This study examined the effect of different therapeutic protocols on macro-mineral profile and reproductive performance of endometritic buffaloes. Endometritic buffaloes were assigned in five treatment and one control groups *viz.* uterine lavage (UL), Levamisole (LEV), PGF_{2α} (PG), Uterine lavage+Levamisole (UL+LEV), Uterine lavage+PGF_{2α} (UL+PG) and control (no therapy) consisting of six buffaloes in each group. The Subclinical endometritic buffaloes were diagnosed using White side test. The screened buffaloes were subjected to different treatment regimen as mentioned earlier. It was observed that the post-treatment calcium and phosphorus level was significantly (P<0.05) higher in all treatment groups as compared to their corresponding pre-treatment values, whereas no significant (P>0.05) change was recorded in control group. The clinical cure rate of endometritis was 33.33, 50.00, 66.67, 66.67, 83.33 and 83.33% in untreated control, UL, LEV, PG, UL+LEV and UL+PG, respectively. The corresponding conception rates were 16.67, 33.33, 50.00, 50.00, 66.67 and 66.67% in untreated control, UL, LEV, PG, UL+LEV and UL+PG, respectively. The results inferred that, uterine lavage along with levamisole or PGF_{2α} can be used effectively to manage endometritic buffaloes. *Key words:* Buffalo, Cloprostenol, Endometritis, Levamisole, Uterine lavage.

How to cite: Singh, N., Singh, B., & Kumar, R. (2021) Therapeutic Outcomes Following Uterine Lavage, Levamisole, PGF2α and their Combinations in Subclinical Endometritic Buffaloes. *The Indian Journal of Animal Patroduction*, 42(2), 17–21, https://doi.org/10.48165/jijar.2021.42.2.3

The Indian Journal of Animal Reproduction, 42(2), 17–21. https://doi.org/10.48165/ijar.2021.42.2.3

INTRODUCTION

Clinical endometritis is defined as purulent uterine discharge detectable in vagina of post-partum cattle or buffaloes 21 days or more postpartum or a mucopurulent discharge detectable in vagina after 26 days postpartum, while, subclinical endometritis is characterized by absence of clinical signs of endometritis (Sheldon *et al.*, 2006). Many studies attempted to associate the presence of bacteria with subclinical endometritis (SCE) which is char-

^{*}Corresponding author.

E-mail address: drrajesh25@gmail.com(Rajesh Kumar)

Recei ed 12-12-2022; Accepted 26-01-2023

Copyright @ Journal of Extension Systems (acspublisher.com/journals/index.php/ijar)

acterized by absence of purulent vaginal discharge with concurrent endometritis based on >5% polymorphonuclear cells (PMNs)/ neutrophils (Pascottini et al., 2020a). Indeed, the role of pathogenic bacteria in genesis of SCE remains to be elucidated (Wagener et al., 2017). Moreover, Wang et al. (2018) stated that SCE is not associated with uterine pathogens. Furthermore, Pascottiniet al. (2020b) reported that uterine microbiota is not different between healthy and SCE cows and the disease is a consequence of dysregulation of inflammation rather than changes in uterine microbiota and advocated that regulation of uterine inflammation is worthy of pursuit for prevention and treatment of SCE. SCE is often undiagnosed (Dutt et al., 2017) resulting in subfertility even after cure. To diagnose the disease, transrectal palpation and ultrasonography of the reproductive tract are commonly undertaken under field conditions (Kasimanickam et al., 2004; Barlund et al., 2008) which are inefficient methods to diagnose SCE.

The ideal therapy for SCE is to eliminate bacterial infection from uterus without compromising uterine defence mechanism (UDM) and has no milk or meat residues. Various drugs has been used to treat uterine infections with variable success (Sheldon et al., 2004; Kumar et al., 2008, 2010; Butani et al., 2009; Makki et al., 2017; Singh et al., 2018; Ahmadi et al., 2019). Levamisole (LEV) is effective in decreasing severity and/or resolving the endometritis and may be used as alternative or adjunct to antibiotic therapy (Singh et al., 2017). Uterine lavage, an important therapeutic tool for the treatment of uterine inflammation eliminates unwanted neutrophils, debris, and other inflammatory products and causes uterine contractions which aid in a physical clearance of uterine contents (Brinsko et al., 2011). Prostaglandins are used to synchronize estrus, induce parturition, and to treat retained fetal membrane, luteinized cyst, pyometra, and chronic endometritis (Weems et al., 2006). The present study was done with hypothesis that uterine lavage, LEV, PG alone or in combinations would ameliorate SCE by augmenting uterine immune system and thereby improving the reproductive outcomes in buffaloes.

MATERIALS AND METHODS

For this study buffaloes were selected from cases presented at Veterinary Clinical Complex, College of Veterinary Science, Acharya Narendra Deva University of Agriculture and Technology (ANDUAT), Kumarganj, Ayodhya and State Veterinary Hospitals in adjoining areas of Kumarganj, Ayodhya, UP (India). This experiment was accomplished under the approval of Institutional Animal Ethics Committee (IAEC), College of Veterinary Science and Animal Husbandry, ANDUAT, Kumarganj, Ayodhya, Uttar Pradesh (India) vide approval number IAEC/CVSc/2019/P-02.

Thirty six buffaloes having problem of repeat breeding with SCE were selected for this study using White side test. Buffaloes were examined 28-35 days in milk (DIM) and selected if they had found positive for white side test (Bhat et al., 2014) without visible muco-purulent vulvar discharge with normal uterus on per rectal palpation. Approximately 6 ml of blood was collected aseptically in clean sterile vials via jugular vein puncture. Samples were centrifuged at 1200g for 15 min at 4°C. The plasma was separated and preserved at -20°C until analysis. The plasma was analyzed for estimation of serum calcium and phosphorus using diagnostic kits as per manufacturer's instructions (ARKRAY Healthcare, Pvt Ltd). Each animal was sampled twice, at start of treatment (pre-treatment) and at subsequent estrus after treatment (post-treatment). At subsequent standing estrus following treatment all the buffaloes were inseminated twice, 12 h apart. If returned to estrus, they were inseminated again at second and third subsequent estrus. Pregnancy was confirmed per-rectally at 45-60 days after last insemination.

At the first examination (28-35 days in milk, DIM) buffaloes with SCE were randomly allocated to six groups: (1) Control group (no treatment); (2) Uterine lavage group (UL): buffaloes were subjected to uterine lavage with 50 mL NSS each time, for six times (total 300mL) as slow intra uterine (IU) infusion at day of estrus; (3) Levamisole group (LEV): endometritic buffaloes were administered with Levamisole (Lemasol-75°, Zydus AH, India) @2.5 mg/kg body weight subcutaneously at 0 (day of estrus), 7th and 14^{th} day of the cycle; (4) PGF₂ α group (PG): buffaloes were administered with Cloprostenol (Vetmate[®], 2 mL vial, Vetcare, India), @ 250 µg intramuscularly on 10th day of the cycle; (5) Uterine lavage+Levamisole group (UL+LEV): the buffaloes were subjected with uterine lavage (same as Group-II) along with Levamisole (same as LEV group); (6) Uterine lavage + PGF₂ α group (UL+PG): the buffaloes were subjected with Uterine lavage (same as Group-II) along with PGF₂ α (same as PG group).

Data were represented as mean \pm Standard error of mean (SEM) and analyzed by using completely randomized design and Duncan's MRT for effect of groups and periods at significance of P value less than or equal to 0.05 (Snedecor and Cochran, 1994).

RESULTS AND DISCUSSION

The results of effect of different treatment on serum calcium of SCE buffaloes have been depicted in Table 1. The

	Calcium	n (mg/dL)	Phosphorus (mg/dL)		
Groups	Pre-treatment	Post-treatment	Pretreatment	Post-treatment	
Control (n=6)	6.51 ± 0.04^{B}	6.59±0.03 ^C	3.57 ± 0.04^{B}	3.66 ± 0.05^{D}	
Uterine Lavage (n=6)	$6.48 {\pm} 0.03^{Ba}$	7.89 ± 0.03^{Bb}	$3.70{\pm}0.04^{Aa}$	4.28 ± 0.05^{Cb}	
Levamisole (n=6)	6.61 ± 0.03^{Aa}	8.11 ± 0.02^{Ab}	$3.73{\pm}0.04^{Aa}$	4.49 ± 0.04^{Bb}	
$PGF_{2\alpha}(n=6)$	6.57±0.03ª	8.04 ± 0.03^{Ab}	$3.65 {\pm} 0.03^{Aa}$	4.38 ± 0.03^{Bb}	
Uterine Lavage +Levamisole (n=6)	$6.51 {\pm} 0.03^{Ba}$	8.17 ± 0.33^{Ab}	$3.71 {\pm} 0.04^{Aa}$	4.62 ± 0.05^{Ab}	
Uterine Lavage + $PGF_{2\alpha}(n=6)$	$6.49 {\pm} 0.04^{Ba}$	8.13 ± 0.03^{Ab}	3.72 ± 0.04^{Aa}	4.57 ± 0.04^{Ab}	

Table 1: Effect	of different treatment	on calcium and	phosphorus st	tatus in	Endometritic	Buffaloes
-----------------	------------------------	----------------	---------------	----------	--------------	-----------

Means with different superscripts within group (a, b) and between groups (A, B, C) differ significantly (P<0.05).

mean values elevated significantly (P<0.05) in treated buffaloes after the treatment than their pre-treatment, whereas, in control group, no significant (P>0.05) change was recorded. At subsequent estrus after treatment, the increase in serum calcium level was highest in UL+LEV group (8.17±0.33mg/dL) followed by UL+PG (8.13±0.03mg/ dL), LEV (8.11±0.02mg/dL), PG (8.04±0.03mg/dL) and UL group (7.89±0.03mg/dL). It can be concluded that among all the treatment groups, UL+ LEV treatment was more effective in restoring the serum calcium level of endometritic buffaloes at subsequent estrus. Our findings were in agreement with the finding of (Sarwar et al., 2002) and (Sharma, 2018), who also reported lower level of calcium in endometritic buffaloes. Low level of calcium in endometritic cows have been reported by many other workers (Kumar et al., 2009; Magnus and Lali, 2009; Jayaram, 2018). The normal reproductive process is compromised in case of severe calcium deficiency (Youngquist and Thrallfall, 2007) possibly due to lack of tone of uterine muscle, as calcium sensitizes the tubular genital tract for action of hormones (Kumar et al., 2020). Majority of infertility problems in cattle (Kumar et al., 2020) and buffaloes (Butani et al., 2011) are due to nutritional deficiency. Plenty of available reports suggest that anestrus, sub-estrus and repeat breeding in cattle (Kumar et al., 2009, 2011) and buffaloes (Butani et al., 2011) might be due to calcium deficiency or imbalance. Furthermore, (Stojikovic et al., 1989) opined that GnRH (Gonadotrophin releasing hormone) induced LH (Luteinizing hormone) release from pituitary is calcium dependent and sub-threshold of calcium leads to failure of LH release.

The results of effect of different treatment on serum phosphorus of SCE buffaloes have been depicted in Table 1. It was observed that the mean values elevated significantly (P<0.05) in treated buffaloes after the treatment than before treatment, whereas, in control no significant change (P>0.05) was recorded. At subsequent estrus after treatment, the increase in serum phosphorous level was highest in UL+ LEV group (4.62 ± 0.05 mg/dL) followed by UL+PG (4.57 ± 0.03 mg/dL), LEV (4.49 ± 0.04 mg/dL), PG (4.38 ± 0.03 mg/dL) and UL group (4.28 ± 0.05 mg/dL). It can be concluded that among all the treatment groups, UL+LEV treatment was more effective in restoring the serum phosphorous level of endometritic buffaloes at subsequent estrus. Our findings were in agreement with the finding of previous reports (Jaychandran *et al.*, 2007; Verma, 2015; Sharma, 2018), who also reported lower level of phosphorous in endometritic cows have been reported by many workers (Chandrahar *et al.*, 2003; Kumar *et al.*, 2009; Pandey *et al.*, 2009; Amle *et al.*, 2014; Jayram, 2018). There is no consensus among scientists that hypophosphatemia is a cause of infertility syndrome in bovines (Noakes *et al.*, 2019).

The overall recovery rate in UL+LEV and UL+PG treated group was recorded as 83.33% and in LEV, PG and uterine lavage treated group the cure rate for SCE was reported as 66.67%, 66.67% and 50%, respectively. The conception rate was also recorded as 66.67% for UL+LEV and UL+PG, 50% for LEV and PG, 33.33% for UL. (Fig 1)



Fig. 1: Effect of different treatment on cure rate and conception rate in Endometritic Buffaloes

Increased conception rate was recorded in all treated buffaloes. Like present findings higher conception rate was also recorded in endometritic cows subjected to uterine lavage (Reddy *et al.*, 2012; Swain *et al.*, 2011), levamisole (Saini *et al.*, 1999; Swain *et al.*, 2011; Biswal *et al.*, 2014; Singh *et al.*, 2017) and $PGF_2\alpha$ (Sood *et al.*, 2003; Sarkar *et al.*, 2006; Biswal *et al.*, 2014, Palanisamy *et al.*, 2014). To the best of our knowledge, no citation available regarding the used treatment, so we could not compare our results with buffaloes.

CONCLUSIONS

It could be concluded that low level of calcium and phosphorus was observed in subclinical endometritis affected buffaloes. Also, the therapeutic management of subclinical endometritis buffaloes with uterine lavage along with either levamisole or PGF_{2α} is better approach with more cure rate as well as resultant higher pregnancy rate. It is advised that the results of the current study need more validation in a larger population due to small sample size.

ACKNOWLEDGEMENTS

The authors are highly thankful to Dean, C.V.Sc. & A.H., ANDUAT, Kumarganj for providing facilities required for research work.

CONFLICT OF INTEREST

None.

REFERENCES

- Ahmadi, M.R., Makki, M., Mirzaei, A., and Gheisari, H.R. (2019). Effects of hypertonic dextrose and paraffin solution as non-antibiotic treatments of clinical endometritis on reproductive performance of high producing dairy cows. *Reprod. Domest. Anim.*, 54(5): 762-771.
- Amle, M., Padodkar, V.P., Shehlar, R. and Birade, H. (2014). Serum biochemical levels of repeat breeder cross bred cows under rural condition of Satara district Maharastra. *Int. J. Adv. Vet. Sci. Tech.*, 3(1): 109-113.
- Barlund, C.S., Carruthers T.D., Waldner C.L., and Palmer C.W. (2008). A comparison of diagnostic techniques for postpartum endometritis in dairy cattle. *Theriogenology.*, **69**: 714-723.
- Bhat, F.A., Bhattacharya, H.K. and Hussain, S.K. (2014). White side Test: A simple and rapid test for evaluation of nonspecific bacterial genital infections of repeat breeding cattle. *Vet. Res. Forum*, **5**(3): 177-180.

- Biswal, S., Das, S., Mohanty, D.N., and Jena, D. (2014). Effect of systemic and local immunomodulation therapies on conception rate in endometritic cows. *Indian J. Field Vets.*, 10(2): 1-4.
- Brinsko, S.P., Blanchard, T.L., Varner, D.D., Schumacher, J., Love, C.C., Hinrichs, K., and Hartman, D.L. (2011). Endometritis. *Manual of Equine Reproduction*, 73-84.
- Butani, M.G., Dhami, A.J. and Kumar, R. (2011). Comparative blood profile of progesterone, metabolites and minerals in anoestrus, suboestrus, repeat breeding and normal cyclic buffaloes. *Indian J. Field Vets.*, 7(2): 20-24.
- Butani, M.G., Dhami, A.J., Kumar, R., Hirani, N.D., Ramani, V.P., and Patel, K.P. (2009). Influence of hormonal and antibiotics therapy on fertility and trace minerals profile in repeat breeding buffaloes. *Indian J. Field Vets.*, 4(3): 12-16.
- Chandrahar, D., Tiwari, P., Awasthi, M.K. and Dutta, G.K. (2003). Serum biochemical profile of repeat breedercows. *Ind. J. Anim. Reprod.*, **24**: 125-127.
- Dutt, R., Singh, G., Singh, M., Sharma, M., Dalal, J., and Chandolia, R.K. (2017). Diagnosis of subclinical endometritis in Murrah buffaloes through Cytobrush technique. *Int. J. Curr. Microbiol. Appl. Sci.*, 6(11): 494-499.
- Jayachandran, S., Selvaraj, P. and Visha, P (2007). Blood biochemical profile in repeat breedingbuffaloes. *Tamilnadu J. Vet. Anim. Sci.*, **3**: 70-73.
- Jayaram, C.R. (2018). Diagnostic and therapeutic methods in postpartum endometritis and its impact on reproductive performance in crossbred cows. Thesis M.V.Sc., Maharashtra Animal and Fishery Sciences University, Nagpur.
- Kasimanickam, R., Duffield, T.F., Foster, R.A., Gartley, C.J., Leslie,
 K.E., Walton, J.S., and Johnson, W.H. (2004). Endometrial
 cytology and ultrasonography for the detection of subclinical endometritis in postpartum dairy cows. *Theriogenology*,
 62: 9-23.
- Kumar, R., Butani, M.G., Dhami, A.J., Kavani, F.S., Bhong, C.D., and Brahmbhatt, M.N. (2008). Isolation and antibiogram of genital microflora from repeat breeding cows and buffaloes. *Indian J. Field Vets.*, 4(2): 5-9.
- Kumar, R., Butani, M.G., Dhami, A.J., Kavani, F.S., Shah, R.G. and Killedar, A. (2011). Management of anoestrus and suboestrus cows using hormonal and nonhormonal drugs. *Indian J. Anim. Reprod.*, **32**(1): 24-27.
- Kumar, R., Dhami, A.J., Butani, M.G., Kavani, F.S., Sarvaiya, N.P., and Killledar, A. (2010). Management of repeat breeding under field condition using hormonal and non hormonal drugs in cows. *Indian J. Anim. Reprod.*, **31**(1): 30-32.
- Kumar, R., Butani, M.G., Dhami, A.J., Kavani, F.S. and Shah, R.G. (2009). Effect of different therapies on fertility and serum

progesterone, metabolites and minerals profile in repeat breeding crossbred cows. *Indian J. Field Vets.*, **5**(2): 1-8.

- Kumar, R., Butani, M.G., Kavani, F.S. and Dhami, A.J. (2020). Hormonal Interventions to Augment Fertility and its Effect on Blood Biochemical Profile in Crossbred Cows. *Haya Saudi J Life Sci.*, 5(9): 176-181.
- Magnus, P. and Lali, K.F. (2009) Serum Biochemical Profile of Post-Partum Metritic Cow. *Vet. World*, **2**(1): 27-28.
- Makki, M., Ahmadi, M.R., Gheisari, H.R., and Nazifi, S. (2017). Cure rate of postpartum endometritis after different treatments in high produce dairy cows. Comp. *Clin. Path.*, **26**: 921-928.
- Noakes, D.E., Parkinson, T.J. and England, G.C.W. (2019). Veterinary Reproduction and Obstetrics 10th (Ed.) Saunders Ltd.
- Palanisamy, M., Napolean, R.E., Selvaraju, M., Krishnakumar, K.,
 Balasubramanian, G.A., Malmarugan, S. and Manokaran,
 S.. (2014). Nature of genital discharge and pH of cervical mucus and uterine flushing before and after treatment in endometritis affected cows. *Int. J. Liv. Res.*, 4(7): 19-24.
- Pandey, V., Singh, A. K., and Sharma, N. (2009). Blood biochemical profile in fertile and repeat breeding crossbred cows under field conditions. *Vet. Pract.*, **10**(1): 112-115.
- Pascottini, O.B., Van, S.S.J., Spricigo, J.F.W., Rousseau, J., Weese, J.S., and LeBlanc, S.J. (2020b). Dynamics of uterine microbiota in postpartum dairy cows with clinical or subclinical endometritic. *Sci. Rep.*, **10**(1):12353.
- Pascottini, O. B. and LeBlanc, S. J. (2020a). Modulation of immune function in the bovine uterus peripartum. *Theriogenology*, **150**:193–200.
- Reddy, N.C.S., Bramhaiah, K.V., Naidu, K.S., Babu, A.J., and Kumar, R.V.S. (2012). Effect of uterine lavage on bacterial count and conception rate in repeat breeder crossbred cows. *Indian J. Anim. Reprod.*, **33**(1): 59-62.
- Saini, P.S., Nanda, A.S., Grewal, A.S. and Singh, J. (1999). Uterine defense modulation for the treatment of repeat breeding due to infectious endometritis in bovines. *Indian J. Anim Sci.*, **69**: 307-309.
- Sarkar, P., Kumar, H., Rawat, M., Varshney, V.P., Goswami, T.K., Yadav, M.C. and Srivastava, S.K.(2006). Effect of administration of garlic extract and $PGF_2\alpha$ on hormonal changes and recovery in endometritis cows. *Asian-Australas J. Anim. Sci.*, **19**(7): 964-969.
- Sarvar, A., Shahid, R.U., Masood, S., Kausar, R. and Shah, S.G. (2002). Serum electrolytes and enzymes in endometritic Nili-Ravi buffaloes of two age groups and at two stages of lactation. *Pak. Vet. J.*, **22**(2); 78-81.
- Sharma (2018). Therapeutic efficacy of ceftiofur sodium, gentamicin sulphate and zingibar officinale extract on recovery of endometritic buffaloes. Thesis MVSc submitted to

Narendra Deva University of Agriculture and Technology, Ayodhya, Uttar Pradesh.

- Sheldon, I. M., Cronin, J. G. and Bromfield, J. J. (2019). Tolerance and Innate Immunity Shape the Development of Postpartum Uterine Disease and the Impact of Endometritis in Dairy Cattle. Annu. Rev. Anim. Biosci., 7: 361–384.
- Sheldon, I.M., Lewis, G.S., LeBlanc, S. and Gilbert, R.O. (2006). Defining post partum uterine disease in cattle. *Theriogenology*, 65: 1516-1530.
- Sheldon, I.M., Noakes, D.E., Rycroft, A.N. and Dobson, H. (2004). Effect of intrauterine administration of oestradiol on postpartum uterine bacterial infection in cattle. *Anim. Reprod. Sci.*, 81: 13-23.
- Singh, B., Gupta, H.P., Prasad, S., and Rajora, V.S. (2018). Effect of Immunomodulators on serum nitric oxide concentration of endometritic repeat breeding crossbred cows. *Indian J. Vet. Res.*, 27(1): 32-35.
- Singh, P.P., Pande, N., Bhavna, and Agrawal, R. (2017). Clinical and biochemical studies on endometritic repeat breeding cows following treatment with levamisol. *Haryana Vet.*, 56(1): 55-57.
- Snedecor, G.W., and Cochran, W.G. (1994). Statistical methods.8th edition, Iowa State University Press, Ames, Iowa, USA.
- Sood, P., Verma, S., Katoch, R.C., Singh, M. and Vaishhta, N.K.. (2003). Impact of uterine microbial panorama on the single injection of PGF₂ alpha in cows with clinical endometritis. *Indian J. Anim. Sci.*, **73**(1): 68-69.
- Swain, P.K., D.N. Mohanty, D.N., Das, S., Barik, A.K., Mishra, P.C., Tripathi, A.K. and Palai, T.K. (2011). Immunomodulation effect of levamisole and immulite on uterine microbial picture in repeat breeding cows. *Indian J. Anim. Reprod.*, **32**(3): 68-70.
- Stojilkovi, S.S., Chang, J.P., Ngo, D., Tasaka, K., Izumi, S. and Catt, K.J. (1989). Mechanism of action of GnRH: the participation of calcium mobilization and activation of protein kinase C in gonadotropin secretion. *J. Steroid Biochem.*, 3(4B): 693-703.
- Verma, S.K. (2015). Study on Incidence of reproductive disorder of cattle and Buffalo in Agro climatic zone of estern Uttar Pradesh. MVSc thesis submitted to CVSc & A.H. N.D.U.A.T. Kumarganj Faizabad.
- Wagener, K., Gabler, C. and Drillich, M. A. (2017). Review of the ongoing discussion about definition, diagnosis and pathomechanism of subclinical endometritis in dairy cows. *Theriogenology*, **94**: 21–30.
- Wang, M.L., Liu, M.C., Xu, J., An, L.G., Wang, J.F. and Zhu, Y.H. (2018). *Front Microbiol.*, 9:2691.
- Youngquist, R.S. and Threlfall, W.R. (2007). Current Therapy in Large Animal Theriogenology. (2nd ed.). Saunders, St. Louis, Missouri.